

## Date: 10/2/14 People in lab: Kelsey Crossen

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**Title:** Chemical Transformation

**Start Time:** 4:35 pm

**Purpose:** Transform successful hmp plasmid to make glycerol stocks and transform ligated product of hmp and pro+RBS to see if it works

**Protocol:** LTM Ed. 2 Chemical Transformations

**Exceptions:**

**Products:**

Sample Label	Description	Source Label	Quantity
CT1 10/2	20 uL	MP1 9/29	1
CT2 10/2	200 uL	MP1 9/29	1
CT3 10/2	20 uL	L3 9/26	1
CT4 10/2	200 uL	L3 9/26	1
CT5 10/2	20 uL	L3 9/26	1
CT6 10/2	200 uL	L3 9/26	1

**Results:**

**Notes: Stop Time:** 8:23 pm

**Next:** Liquid cultures of any growth for minipreps

## Date: 10/2/14 People in lab: Kira Buckowing

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**Title:** Digest and ligation of norV and psB1C3 backbone; Prep of hmp plasmids for cDNA lab sequencing

**Start Time:** 4:50 pm

**Purpose:** To see if the hmp product is correct and to see if the sample of norV from 11/25/13 are correct and can be used at all

**Protocol:** LTM Ed. 2 Digestion and Ligation and Vanessa's instructions for cDNA

**Exceptions:**

**Products:**

Sample Label	Description	Source Label	Quantity
L1 10/2	Ligation of A1 and backbone	A1 11/25/13	1
L2 10/2	Ligation of norV and backbone	A2 11/25/13	1

**Results:**

**Notes:** cDNA products stored in the freezer for sequencing

**Stop Time:** 7:30 pm

**Next:** Transformation of ligations to see if anything grows and sequencing

## Date: 10/3/14 People in lab: Kelsey Crossen

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**Title:** Broth cultures of CT1-6 10/2

**Start Time:** 2 pm

**Purpose:** Miniprep preparation

**Protocol:** LTM Ed. 2 Inoculation

**Exceptions:**

**Products:**

Sample Label	Description	Source Label	Quantity
BC1 10/3	hmp psB1C3	CT2 10/2	1
BC2 10/3	hmp psB1C3	CT1 10/2	1
BC3 10/3	hmp psB1C3	CT1 10/2	1
BC4 10/3	hmp with pro+RBS	CT1 10/3	1
BC5 10/3	hmp with pro+RBS	CT3 10/2	1
BC6 10/3	hmp with pro+RBS	CT3 10/2	1
BC7 10/3	hmp with pro+RBS	CT4 10/2	1
BC8 10/3	hmp with pro+RBS	CT4 10/2	1
BC9 10/3	hmp with pro+RBS	CT5 10/2	1
BC10 10/3	hmp with pro+RBS	CT5 10/2	1
BC11 10/3	hmp with pro+RBS	CT6 10/2	1
BC12 10/3	hmp with pro+RBS	CT6 10/2	1

**Results:**

**Notes:**

**Stop Time:** 2:45 pm

**Next:** Glycerol stocks of BC1-3 and miniprep of the rest

## Date: 10/3/14 People in lab: Kira Buckowing

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**Title:** Chemical Transformation of L1 and L2 10/2

**Start Time:** 8:30 pm

**Purpose:** To test if the digestion and ligation of norV was successful

**Protocol:** LTM Ed. 2 Chemical Transformation

**Exceptions:**

**Products:**

Sample Label	Description	Source Label	Quantity
CT1 10/3	20 uL	L1 10/2	1
CT2 10/3	200 uL	L1 10/2	1
CT3 10/3	20 uL	L2 10/2	1
CT4 10/3	200 uL	L2 10/2	1

**Results:**

**Notes:** Wrong plates used by mistake at the last step and the procedure was started over and redone (original start time ~5 pm)

**Stop Time:** 11:30 pm

**Next:** Inoculation of any colonies that grow

## Date: 10/4/14 People in lab: Kelsey Crossen

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**Title:** Minipreps

**Start Time:** 2:50 pm

**Purpose:** Isolate igem Plasmid cocntaining hmp and a promoter+RBS site

**Protocol:** LTM Ed. 2 Miniprep

**Exceptions:**

**Products:**

Sample Label	Description	Source Label	Quantity
MP1 10/4	ng/uL = 210.9; 260/280 = 2; 260/230 = 2.1	BC5 10/3	1
MP2 10/4	ng/uL = 144.7; 260/280 = 2.12; 260/230 = 2.23	BC6 10/3	1
MP3 10/4	ng/uL = 446.8; 260/280 = 1.9; 260/230 = 1.61	BC7 10/3	1
MP4 10/4	ng/uL = 173.4; 260/280 = 2; 260/230 = 2.03	BC8 10/3	1
MP5 10/4	ng/uL = 144.8; 260/280 = 1.97; 260/230 = 1.68	BC9 10/3	1
MP6 10/4	ng/uL = 234.5; 260/280 = 2.02; 260/230 = 2.17	BC10 10/3	1
MP7 10/4	ng/uL = 314.3; 260/280 = 1.9; 260/230 = 2.21	BC11 10/3	1
MP8 10/4	ng/uL = 195.9; 260/280 = 1.92; 260/230 = 2.19	BC12 10/3	1

**Results:**

**Notes:**

**Stop Time:** 4:55 pm

**Next:** Digest with P and X and run gel to check for insert and vector

## Date: 10/4/14 People in lab: Kira Buckowing

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**Title:** Innoculation from CT1-2 10/2

**Start Time:** 3:15 pm

**Purpose:** To get more liquid cultures of hmp in psB1C3

**Protocol:** LTM Ed. 2 Innoculation

**Exceptions:**

**Products:**

Sample Label	Description	Source Label	Quantity
BC1 10/4	hmp	CT1 10/2	1
BC2 10/4	hmp	CT1 10/2	1
BC3 10/4	hmp	CT2 10/2	1
BC4 10/4	hmp	CT2 10/2	1

**Results:**

**Notes:**

**Stop Time:** 3:30 pm

**Next:** Glycerol stocks of one and minipreps of the other 3

## Date: 10/5/14 People in lab: Kira Buckowing

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**Title:** Glycerol stock of hmp

**Start Time:** 12 pm

**Purpose:** To have back up stock of the proper hmp plasmid

**Protocol:** LTM Ed. 2 Stocks

**Exceptions:**

**Products:**

Sample Label	Description	Source Label	Quantity
hmp 1 Glycerrstock 10/5	hmp glycerol stock	BC2 10/4	1
hmp 2 Glycerstock 10/5	hmp glycerol stock	BC2 10/4	1
hmp 3 Glycerstock 10/5	hmp glycerol stock	BC2 10/4	1

**Results:**

**Notes:** Used Dr. Westenberg's 40% Glycero

**Stop Time:** 12:30 pm

**Next:** Minipreps of the other three inoculations

## Date: 10/6/14 People in lab: Kira Buckowing

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**Title:** Minipreps of hmp BCs

**Start Time:** 9:30 am

**Purpose:** To isolate the plasmid with hmp in the standard shipping backbone

**Protocol:** LTM Ed. 2 Miniprep

**Exceptions:**

**Products:**

Sample Label	Description	Source Label	Quantity
MP1 10/6	hmp	BC1 10/4	1
MP2 10/6	hmp	BC3 10/4	1
MP3 10/6	hmp	BC4 10/4	1

**Results:**

**Notes:**

**Stop Time:** 10:30 am

**Next:** Nanodrop for concentrations and sequencing again