Name: Katie Fitton

Date: 7/22/19

WEEKLY ONE SENTENCER:

Continue testing RBS primers & promoters

DAILY ONE SENTENCER:

clone RBS primers into mcherry

METHOD(S) OF CHOICE:

PCR
PCR purification
gel electrophoresis

EXPECTED RESULT:

band @ 1 kb

(☒ check if APE file exists on Drive)

TEMPLATE(S): ~~@~~ mcherry

OLIGO(S): 74-78, 363

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE: 362

NEGATIVE:

w/o polymerase

CONFIRMATION?

fwd gel

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Jennifer Zhang

Date: 7/24/14

WEEKLY ONE SENTENCER: continue troubleshooting RBS and promoter primers

DAILY ONE SENTENCER: grow up colonies from transformations practice group presentation

METHOD(S) OF CHOICE:
cultures (LB + CM)

EXPECTED RESULT:
cultures to be miniprepped tomorrow

(☐ check if APE file exists on Drive)

TEMPLATE(S):

OLIGO(S):

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE:

NEGATIVE:

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Jennifer Zhang

Date: 7/25/14

WEEKLY ONE SENTENCER: continue troubleshooting RBS and promoter pairs

DAILY ONE SENTENCER: - miniprep RH2, RH1 then send for sequencing
promoter PCR w/ RBS
digest/ligate then transform

METHOD(S) OF CHOICE:
miniprep
PCR
digest / ligate
transformation

EXPECTED RESULT:
colonies on LB + CM plates
band ~ 1kb

(☒ check if APE file exists on Drive)

TEMPLATE(S): RBS RH2

OLIGO(S): 80 - 85, 363

THERMOCYCLER SETTINGS: 94 - 2' [94' - 15", 60 - 15", 68 - 1.5'] x 31
68 C - 4'; 4°C

CONTROLS:

POSITIVE:
362

NEGATIVE: no template

CONFIRMATION?
gel

GEL LANE ORDER:

1 kb	PH1	PL1	PH1	PL1	RH1	PL1
	RH2	RH2	RH1	RH1	RL1	RL1

GEL PICTURE / OTHER SPACE:

Name: Troy von Beck

Date: 8/5/2014

WEEKLY ONE SENTENCER:

Finish formatting the wiks Notebook

DAILY ONE SENTENCER:

Transform selected promoters from Biobrick plates

METHOD(S) OF CHOICE:

Heat Shock
Cm Antibiotic selection

EXPECTED RESULT:

Colonies with useful Biobricks

(☐ check if APE file exists on Drive)

TEMPLATE(S): To be determined

OLIGO(S):

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE:

NEGATIVE:

DH5α untransformed

CONFIRMATION?

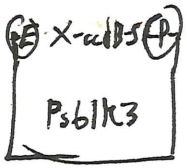
GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

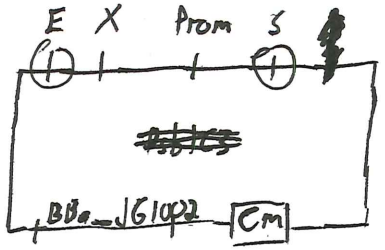
Which Promoter?

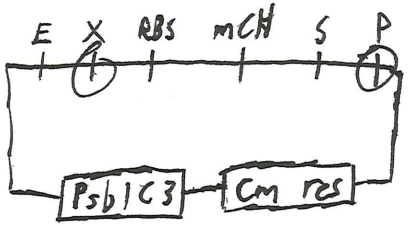
Anderson Promoters

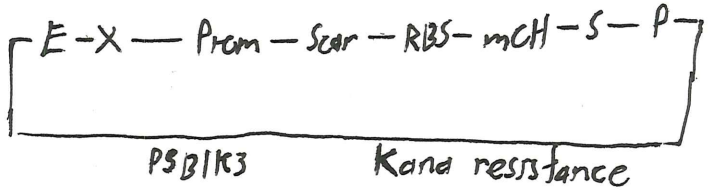
Part #	Strength	Location
BBa_J23100	1	2014 kit Plate 4, well 17D
BBa_J23102	.86	" " , well 17H
BBa_J23101	.7	" " , well 17F



BBa_J04450







Name: Nishu. C

Date: 8/6/14

WEEKLY ONE SENTENCER:

- Work on cloning RBS into α, β & γ
- Cloning RBS into sumo 3

DAILY ONE SENTENCER:

- Clone sumoB, sumoC, α-1, β & γ into bioBrick plasmid

METHOD(S) OF CHOICE:

- digestion (E&P), ligation, Miniprep gel, gel extraction & purification

EXPECTED RESULT:

Bands @ 1600 bp for α-1 & 554 bp for γ subunit
467 bp " sumoB
1088 bp " sumoC
1211 bp " β subunit

Also transformed colonies on Cm plates

(☑ check if APE file exists on Drive)

TEMPLATE(S): mCherry plasmid

OLIGO(S): N/A

THERMOCYCLER SETTINGS: Digest 37 (1 hr @ 37°C)

CONTROLS:

POSITIVE:

NEGATIVE: H_2O without inserts. w/ vector

CONFIRMATION?

Miniprep gel

GEL LANE ORDER:

1 to 6

GEL PICTURE / OTHER SPACE:

Name: Jennifer Zhang

Date: 8/11/14

WEEKLY ONE SENTENCER: put proteins B,C in biobrick format ✓

DAILY ONE SENTENCER: ~~digest~~ protein B,C PCR product, ligate with PSB-1C3 backbone, transform ✓

METHOD(S) OF CHOICE:
digest/ligate w/ E/P
PCR purify
gel extract
~~transformation~~ ✓

EXPECTED RESULT:
α → 1.5 kb
mcherry → 2 kb ✓

~~X~~ (☐ check if APE file exists on Drive)

TEMPLATE(S): genes α, β, γ, B, C ✓

~~OLIGO(S):~~

THERMOCYCLER SETTINGS: 37°C 1 hr
80°C 20 min ✓

CONTROLS:

POSITIVE:

NEGATIVE:

CONFIRMATION?
gel

GEL LANE ORDER:
1kb α mch ✓

GEL PICTURE / OTHER SPACE:

Name: Aniruddh Joshi

Date: 8/12/14

WEEKLY ONE SENTENCER:

Inserting genes α, β, γ, B and C into pSB-1C3

DAILY ONE SENTENCER:

Digest genes α, β, γ, B, C and PSB-1C3 backbone

METHOD(S) OF CHOICE:

EXPECTED RESULT:

(☐ check if APE file exists on Drive)

TEMPLATE(S):

OLIGO(S):

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE:

NEGATIVE:

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Aniruddh Joshi

Date: 8/14/14

WEEKLY ONE SENTENCER: inserting α, β, γ, B and C into psB-1C3 backbone

DAILY ONE SENTENCER: Miniprep cultures of α, β, γ, B, C
Digesting genes α, β, γ, B and C and ligate into mCherry

METHOD(S) OF CHOICE:
Digestion (E & P) ✓
ligation (instant)
Transformation

EXPECTED RESULT:
-Bands @ 1600 bp for α, 1211 for β
~~1140~~ for mC 554 for γ
467 ~~467~~ for B
1088 for C ✓

(☑ check if APE file exists on Drive)

TEMPLATE(S): mCherry ✓

OLIGO(S):

THERMOCYCLER SETTINGS: Digest with 80°C @ 20 min

CONTROLS: N/A

POSITIVE: ✓

NEGATIVE: N/A

CONFIRMATION?
Gel

GEL LANE ORDER: ✓
1 kb α β γ B C mC

GEL PICTURE / OTHER SPACE:

Contamination suspected in water or PE buffer ✓
*plan to replace both

Name: Jennifer Zhang

Date: 6/14/14

WEEKLY ONE SENTENCER: continue trying to put genes α, β, γ, B, C into PSB-1C3

DAILY ONE SENTENCER: digest α, β, γ, B, C, mCherry @ E/P ligating and transforming miniprep β, γ, B, C

METHOD(S) OF CHOICE:
digest E/P
gel extraction
ligation
transformation

EXPECTED RESULT:
α → 1600 bp C → 1088
β → 1200
γ → 558
B → 467
mCh → 2kb

(☒ check if APE file exists on Drive)

TEMPLATE(S): α, β, γ, B, C, mCherry

~~OLIGO(S):~~

THERMOCYCLER SETTINGS: 37°C @ 1hr
60°C @ 20 min

CONTROLS:

POSITIVE:

N/A

NEGATIVE:

N/A

CONFIRMATION?

gel

GEL LANE ORDER:

1kb α β 1 B C mC

GEL PICTURE / OTHER SPACE:

contamination → threw out products

Name: Mishi.C

Date: 8/14/14

WEEKLY ONE SENTENCER:
-Continue trying to insert all genes into the psB1C3

DAILY ONE SENTENCER:
-Miniprep cultures of α, B, C, β ✓
-Digest, ligate & Transform genes

METHOD(S) OF CHOICE:
-Digestion (E&P) -Gel extraction + purification
-ligation (Instant) ✓
-Transformation

EXPECTED RESULT:
-bands @ 1600 bp for α 1211 for β ! Transformed colonies
2200 for mC 554 for γ !
967 for B !
1088 for C ✓

(☑ check if APE file exists on Drive)

TEMPLATE(S): mCherry

OLIGO(S):

THERMOCYCLER SETTINGS: Digest with 80°C @ 20 min ✓

CONTROLS: N/A

POSITIVE:

NEGATIVE: N/A

CONFIRMATION?
Gel

GEL LANE ORDER:
1kb α β γ B S mC

GEL PICTURE / OTHER SPACE: How did this turn out?

Name: Troy

Date: 8/14/2014

WEEKLY ONE SENTENCER:

Ligate iGEM Anderson Promoters with our RBS mcherry

DAILY ONE SENTENCER:

" " "

METHOD(S) OF CHOICE:

Digest RH2 mCherry X+P

Digest BBa_J23 (100-102) S+P

~~Big~~ gel extract RH2 mcherry insert + BBa_J2310N Backbones

EXPECTED RESULT: ~~Ligate insert w/ backbones.~~

~~Transform + Plate on Kan~~ Amp plates

Pink colonies tomorrow

(☐ check if APE file exists on Drive)

TEMPLATE(S): RH2 mCherry, BBa_J23100, BBa_J2310, BBa_J23102

OLIGO(S): X, S, P

THERMOCYCLER SETTINGS: Digest

CONTROLS: None

POSITIVE: None

NEGATIVE: Control ~~Kan~~ Amp Plate

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Poor Quality Gel Extraction

Low Yields w/ contamination, did not progress to ligation

Name: Coleen Tran

Date: 8/14/14

WEEKLY ONE SENTENCER:

working on promoter + RBS + mCherry ✓

DAILY ONE SENTENCER:

confirming sequencing & helping Jennifer + Tishni

METHOD(S) OF CHOICE:

- PCR
- digest + ligate ✓

EXPECTED RESULT:

(beta) B: ~1200 bp
C: ~1100 bp
B: 450

γ: 550

α: 1600 ✓

(☐ check if APE file exists on Drive)

TEMPLATE(S): ammo genes

OLIGO(S): 90, 91

THERMOCYCLER SETTINGS: 98°C - 30" - { 98°C - 10" ; 60°C - 20" ; 72°C - 1' } x31 ; 72° - 2' ; 4°C ✓

CONTROLS:

POSITIVE:

NEGATIVE:

no template

CONFIRMATION?

gel

GEL LANE ORDER:

L B C β γ α N ✓

GEL PICTURE / OTHER SPACE:

↳ results?

How did this turn out?

Name: Stefan Tassoulas

Date: 08/5/2014

WEEKLY ONE SENTENCER:
- Clone RBS into αBγ
- Finish collaboration with GS. ✓

DAILY ONE SENTENCER:
- Transform αBγ into psB1C3 DH5α ✓
- Work with G.S.

METHOD(S) OF CHOICE:
- Digestion, PCR machine, Ligation
- E+P ✓

EXPECTED RESULT:
Transformed αBγ clones ✓

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

(☐ check if APE file exists on Drive)

TEMPLATE(S): ~~Kan~~ pU57-b, pU57-γ, TOPO-α

OLIGO(S): N/A

THERMOCYCLER SETTINGS: Digestion protocol ~~37° for 2 hrs, 65° 20min, 4°/6° forever~~
37°C for 1hr, 80°C for 20mins, 4°C forever

CONTROLS:

POSITIVE:

NEGATIVE:
digest Neg control
H_2O + cutsmart + E+P

Name: Coleen Tran

Date: 8/5/14

WEEKLY ONE SENTENCER:

working on promoter + RBS + mCherry clones

DAILY ONE SENTENCER:

- PCR promoter primer into RBS + mCherry clones
- contact Tom Muir from UVA about collab.

METHOD(S) OF CHOICE:

- PCR
- gel

EXPECTED RESULT:

~1 kb

CONFIRMATION?

gel

GEL LANE ORDER:

L H86 H87 H88 _ L86 L87 L88 WT

GEL PICTURE / OTHER SPACE:

140805 Promoter + RBS + mCherry

(☐ check if APE file exists on Drive)

TEMPLATE(S): [redacted] #2, [redacted] L1

OLIGO(S): [redacted], 363, 86 - 88

THERMOCYCLER SETTINGS:

CONTROLS: 98°C - 30" { 98°C - 10", 60°C - 20", 72°C - 20" } x31 ; 72 - 2', 4°C

POSITIVE:

3682

NEGATIVE: no template ✓

Name: Katie Fitton

Date: 8/5/14

WEEKLY ONE SENTENCER:

Work on putting sMMO genes in biobrick format

DAILY ONE SENTENCER:

BAC

transform sMMO genes w/ biobrick plasmid

METHOD(S) OF CHOICE:

digestion (E & P), gel extract, ligation, transformation

EXPECTED RESULT:

α - 1600 bp

B & β - 1500 bp

γ & C - 1600 bp

(☐ check if APE file exists on Drive)

TEMPLATE(S): mcherry plasmid

OLIGO(S):

THERMOCYCLER SETTINGS: 1 hour @ 37°C

CONTROLS:

POSITIVE:

NEGATIVE:

w/o restriction enzymes (E & P)

CONFIRMATION?

thin gel of just plasmid before digestion

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Nishi.C

Date: 8/5/14

WEEKLY ONE SENTENCER:

- Working on cloning RBS into α, β, γ
- Cloning RBS into 3H SUMO's

DAILY ONE SENTENCER:

- Cloning Sumo B, SumoC, α-1, β, γ into biobrick plasmid.

METHOD(S) OF CHOICE:

- Digestion, Ligation with (E&P) &
  - Using Digest 37

EXPECTED RESULT:

Bands @: 1600 bp for α-1
467 bp " sumo B
1098 bp "SumoC
1211 bp for β subunit
554 bp for γ subunit

1 Transformed clones

(☐ check if APE file exists on Drive)

TEMPLATE(S): pU 57-β, pU 57-γ, Topo - α

OLIGO(S): N/A

THERMOCYCLER SETTINGS: Digest 37

CONTROLS:

POSITIVE: N/A

NEGATIVE: Digested Neg control of H_2O + cutsmart + EP

CONFIRMATION?

Gel

GEL LANE ORDER:

1kb β γ α-1 C B

GEL PICTURE / OTHER SPACE:

Name: Eileen Tran

Date: 8/6/14

WEEKLY ONE SENTENCER:

working on promoter + RBS + mCherry

DAILY ONE SENTENCER:

digest + ligate promoter + RBS + mCherry into vector

METHOD(S) OF CHOICE:

digest, ligate, transformation

EXPECTED RESULT:

vector: ~2 kb

insert: ~1 kb

(☐ check if APE file exists on Drive)

TEMPLATE(S): 486-88, L86-88

OLIGO(S):

THERMOCYCLER SETTINGS: 37°C for 1 hours (digestion)

CONTROLS:

POSITIVE:

NEGATIVE: no template

CONFIRMATION?

Do agar plates

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Katie Fitton

Date: 8/26/14

CONFIRMATION?

~~PCR~~ gel

WEEKLY ONE SENTENCER:

Work on plating sMMO genes (B&C) in biobrick format

GEL LANE ORDER:

DAILY ONE SENTENCER:

~~grow vects & colonies~~ transform sMMO genes w/ biobrick plasmid again

GEL PICTURE / OTHER SPACE:

METHOD(S) OF CHOICE:

digestion (E & P), gel extract, ligation, & transformation

EXPECTED RESULT:

~~cell~~ ~~[illegible]~~ ~~[illegible]~~

467 bp sMMO B

1028 bp sMMO C

~~[illegible] bp~~

(☐ check if APE file exists on Drive)

TEMPLATE(S): mcherry plasmid

OLIGO(S): N/A

THERMOCYCLER SETTINGS: ~~Digest~~ 1 hour @ 37°C

CONTROLS:

POSITIVE:

NEGATIVE:

H_2O instead of DNA w/ vector

Name: Rishi.C

Date: 8/7/14

WEEKLY ONE SENTENCER:
- Work on cloning RBS into α, β, ε, & γ α, β, γ ✓
  - Cloning RBS into SMMO's

DAILY ONE SENTENCER:
- PCR of all the subunits with Q5 (α-1, β, B, C, γ)!
- Digestion of all genes (E&P)
- Digestion with just vectors (RH2 & mCherry) (E&P)!
- Dephosphorylation for the vectors (new method)!

METHOD(S) OF CHOICE:
- PCR set in thermocycler
- Digestion (E&P)
- Ligation (overnight for PCR stuff)
- gel extraction & purification
- Dephosphorylation

EXPECTED RESULT:
For gel: bands @ 1600bp for α-1
467 for smmoB
1088 for " C
1211 " " β subunit
& 554 bp for γ
& transformation of All gene digestion

(☑ check if APE file exists on Drive)

TEMPLATE(S): mCherry plasmid ! All the genes for Q5 PCR !

OLIGO(S): 90 & 91 ✓

THERMOCYCLER SETTINGS: Q5 ST

CONTROLS:

POSITIVE:

NEGATIVE: No template. ✓

CONFIRMATION?
Gel ✓

GEL LANE ORDER:
1Kb

GEL PICTURE / OTHER SPACE:

C: 3.63 μL insert
1.37 μL vector

γ: 3.19 μL insert
1.81 μL Vector

β: 3.59 μL insert
1.41 μL vector

B: 2.16 μL insert
2.84 μL vector

Name: Coleen Tran

Date: 8/7/14

WEEKLY ONE SENTENCER:

working on ~~RBS~~ Bba promoter + RBS+mCherry ✓

DAILY ONE SENTENCER:

miniprepping Tracy's RFP – Andersen promoter cell cultures ; culturing promoter + RBS+mCherry clones

METHOD(S) OF CHOICE:

- Miniprep ✓
- cell culturing ✓

EXPECTED RESULT:

- pink cells ✓

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

(☐ check if APE file exists on Drive)

TEMPLATE(S): BBa_J23100-02 ; RHI RHI RHI RLI RLI RLI

OLIGO(S): PHI PH2 PLI PHI ~~PH2~~ PLI

THERMOCYCLER SETTINGS:

CONTROLS: ✓

POSITIVE:

NEGATIVE:

Name: Coleen Tran

Date: 8/8/14

WEEKLY ONE SENTENCER:

working on RPS + promoter + mCherry

DAILY ONE SENTENCER:

miniprep promoter + RPS + mCherry culture

METHOD(S) OF CHOICE:

miniprep

EXPECTED RESULT:

pink colonies

(☐ check if APE file exists on Drive)

TEMPLATE(S):

OLIGO(S):

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE:

NEGATIVE:

CONFIRMATION?

sequencing

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Nidhi. C

Date: 8/8/14

WEEKLY ONE SENTENCER:

- Continue cloning ~~RBS into~~ genes into psB1C3

DAILY ONE SENTENCER:

- Digest & ligate all genes, transform all genes into psB1C3

METHOD(S) OF CHOICE:

- Digestion
- Ligation
- Transformation

EXPECTED RESULT:

- Transformed colonies of all genes.

(☑ check if APE file exists on Drive)

TEMPLATE(S):

OLIGO(S):

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE:

NEGATIVE:

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Troy

Date: 8/12/2014

WEEKLY ONE SENTENCER:

Ligate iGEM Anderson Promoters with our RBS mcherry

DAILY ONE SENTENCER:

" " "

METHOD(S) OF CHOICE:

Digest RH2 mCh E+X
Digest BBa_J2310N E+S
Gel extract RH2 mCh Backbone + BBa_J2310N inserts

EXPECTED RESULT: ~~Ligate inserts w/ Backbone transform and plate on Cm plates~~

inserts were to small to bind to the spin column, no DNA was retained for ligations

Pink colonies tomorrow

(☐ check if APE file exists on Drive)

TEMPLATE(S): RH2/RL1 mCh, BBa_J23100, BBa_J23101, BBa_J23102

OLIGO(S): E, X, S

THERMOCYCLER SETTINGS: Digest

CONTROLS: None

POSITIVE: None

NEGATIVE: Control Cm Plate

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Troy

Date: 8/13/2014

WEEKLY ONE SENTENCER:

Ligate iGEM Anderson Promoters with our RBS mCherry.

DAILY ONE SENTENCER:

" " "

METHOD(S) OF CHOICE:

Digest Alt2 mCh X+P
Digest BBa_J23100 S+P
Gel extract Alt2 mCh insert + BBa_J23100 Backbones

EXPECTED RESULT:

Ligate insert w/ Backbones
Transform + Plate on Amp plates

Pink colonies tomorrow

(☐ check if APE file exists on Drive)

TEMPLATE(S): Alt2 mCherry, BBa_J23100, BBa_J23101, BBa_J23102

OLIGO(S): X, S, P

THERMOCYCLER SETTINGS: Digest

CONTROLS: None

POSITIVE: None

NEGATIVE: Control Amp Plate

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Update: Colonies did not grow on Antibiotic Plates

Name: Troy

Date: 8/15/2014

CONFIRMATION?

WEEKLY ONE SENTENCER:

Ligate existing iGEM promoters with our RBS mCherry.

GEL LANE ORDER:

DAILY ONE SENTENCER:

Resupply ces with Buffers, plates, autoclaved materials;

GEL PICTURE / OTHER SPACE:

METHOD(S) OF CHOICE:

Recipes

gel extraction from before came out poorly. Continue next week!

EXPECTED RESULT:

PB Buffer Autoclaved tips + tubes
PE Buffer Clean fridge + freezer Boxes
Lm Plates

(☐ check if APE file exists on Drive)

TEMPLATE(S):

OLIGO(S):

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE:

NEGATIVE:

Name: Colem Tuson

Date: 8/15/14

WEEKLY ONE SENTENCER:

working on prom. + RBS + mCherry

DAILY ONE SENTENCER:

working on sMMP9 gene; cloning into PSB1C3 ✓

METHOD(S) OF CHOICE:

- PCR — ligate
- digestion —

β, γ

EXPECTED RESULT:

α: 1000 γ: 550

β: 1200 B: 450

C: 1100

(☐ check if APE file exists on Drive)

TEMPLATE(S):

OLIGO(S):

THERMOCYCLER SETTINGS: 98°C - 30" { 98° - 10" ; 68°C - 20" ; 72°C - 1' } x 34 ; 72° - 2' ; 4°C

CONTROLS:

POSITIVE:

NEGATIVE:

no template

CONFIRMATION?

gel

GEL LANE ORDER:

L B C β γ α N

GEL PICTURE / OTHER SPACE:

Name: Anirudh Joshi

Date: 8/15/14

WEEKLY ONE SENTENCER:

~~Inserting α, β, δ, B, C genes into pSB-1C3 backbone~~

DAILY ONE SENTENCER:

~~Add~~ Making Thin and Thick gels

METHOD(S) OF CHOICE:

Gel Making

EXPECTED RESULT:

Gel

(☐ check if APE file exists on Drive)

TEMPLATE(S):

OLIGO(S):

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE:

NEGATIVE:

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

* β, γ into pSB1C3 for next week and grow cultures of α, B, C

Name: Jennifer Zhang

Date: 8/15/14

WEEKLY ONE SENTENCER: ~~continue putting genes α, β, γ, B, C in pSB-1C3 backbone~~

DAILY ONE SENTENCER: make thin and thick gels

METHOD(S) OF CHOICE:

~~*~~ β, γ SE PUT into pSB1C3

EXPECTED RESULT:

(☐ check if APE file exists on Drive)

TEMPLATE(S):

OLIGO(S):

THERMOCYCLER SETTINGS:

CONTROLS:

POSITIVE:

NEGATIVE:

CONFIRMATION?

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Colem Tran

Date: 8/19/14

WEEKLY ONE SENTENCER:

working on promoter + RBS + mCherry

DAILY ONE SENTENCER:

cloning β + α into pSB1c3 vector

METHOD(S) OF CHOICE:

digest E+P, ligate, transformation

EXPECTED RESULT:

α - 1600 bp

β - 1200 bp

mCherry backbone - 2 kb.

(☐ check if APE file exists on Drive)

TEMPLATE(S): α, β, mCherry

OLIGO(S):

THERMOCYCLER SETTINGS: 37°C - 10' ; 80°C - 20', 4°C

CONTROLS:

POSITIVE:

NEGATIVE:

CONFIRMATION?

colonies on plates

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE: