Name: Coleen Tran

Date: 6/17/14

WEEKLY ONE SENTENCER:
redoing / characterizing RBS + promoter in mCherry

DAILY ONE SENTENCER:
all RBS + P_{Hz}

METHOD(S) OF CHOICE:
PCR ⇒ gel extract ⇒ digest ⇒ ligate

EXPECTED RESULT:
> 1000 bp ✓

(☑ check if APE file exists on Drive)

TEMPLATE(S): mCherry + RBS (which? H_1-L_1?), ~~P_{Hz} (81)~~

OLIGO(S): P_{Hz} (81) ✓, 373 ✓

THERMOCYCLER SETTINGS: 94-2'; {94'-15"; 65-15"; 68-1.5'} × 31 68°C-4'; 4°C ✓

CONTROLS:

POSITIVE:

template only? now.... no + needed

NEGATIVE:

no polymerase

CONFIRMATION?
gel

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Name: Troy von Beck

Date: 6/17/2014

WEEKLY ONE SENTENCER: Finish initial wiki HTML code, begin more advanced/better looking formatting.

DAILY ONE SENTENCER: Gel Extract Re-PCR products from Yesterday. Create all accessory pages and improve the page links.

METHOD(S) OF CHOICE:

EXPECTED RESULT:
9 Bands of mCherry ~1100 bp each with an RBS + Promoter combo
(unique)

(☒ check if APE file exists on Drive)

TEMPLATE(S): 9 Bands extracted on Friday

OLIGO(S): Same Oligos used on each respective Band Previously

THERMOCYCLER SETTINGS: Same as on Friday

CONTROLS: NTC

POSITIVE:
P??/R??/mCherry/VR

NEGATIVE:
NTC

CONFIRMATION?
Gel Electrophoresis

GEL LANE ORDER:

GEL PICTURE / OTHER SPACE:

Results: Successfully extracted bands for 2 of the 9 Promoter/RBS combos PH2/RL1 & PH1/RM1
39 ng/uL
57 ng/uL

Name: Tishi Choudhury

Date: 06/17/19

WEEKLY ONE SENTENCER:
Complete ~~set~~ the RBS primer + Promoter primer addition
Finish crowdfunding things.

DAILY ONE SENTENCER:
Digest PCR amplification if gel shows up nicely. Also add in the promoter to RBS primer & PCR

METHOD(S) OF CHOICE:
- PCR, gel extraction & purification (& Prom L1)
- Digestion & ligation of amplified the promoters & RBS; + Transformation

EXPECTED RESULT:
Good concentrations on nanodrop for the Promoter addition
Also bands ~1000 bp

CONFIRMATION?
Running a gel & finding ~1000bp bands ✓

GEL LANE ORDER:
For mCherry + RBS + Prom: 1kb B1 B2 B3 B4 B5 B6 → Ei1 no dig.

GEL PICTURE / OTHER SPACE:

(☑ check if APE file exists on Drive) ✓

TEMPLATE(S): RBS + H1, H2, M1, M2 & L1 inserted in front of mCherry.

OLIGO(S): 273, PL1

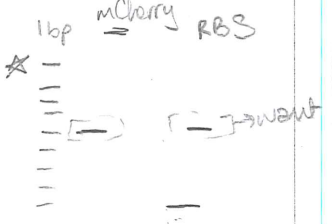
THERMOCYCLER SETTINGS: 65°? 94-2', {94'-15", 65-15", 68°C-1.5'}$^{\times 31}$ – 68°C-4', 4°C forever ✓

CONTROLS:

POSITIVE: With polymerase ☺ ✓

NEGATIVE: No polymerase ✓

★ Did ligation mCherry with RBS & the mCherry backbone
↳ ~~Normal~~ ↳ Transformed using Dwight & our comp. cells.
☺
The end.



★ Did PCR for 35 combinations.

Primers \ Template	RH1	RH2	RM1	RM2	RL1
PH1	A1	B1	C1	D1	E1
PH2	A2	B2	C2	D2	E2
PM1	A3	B3	C3	D3	E3
PM2	A4	B4	C4	D4	E4
PL1	A5	B5	C5	D5	E5
PL2	A6	B6	C6	D6	E6

for negatives: A1 – E1 with no polymerase and primer PH1.

~~★ Adding in ligation.~~

Name: Stefan Tassoulas

Date: 06 / 17 / 14

WEEKLY ONE SENTENCER:
Characterize rbs & promoter pairs in E. coli:
Work on funding (Macrogen) →

DAILY ONE SENTENCER:
Insert Promoter in rbs mcherry insert

METHOD(S) OF CHOICE:
PCR
Gel extraction

EXPECTED RESULT:
~979 bp promoter-rbs-mCherry ✓

(☐ check if APE file exists on Drive)

TEMPLATE(S): RBS(H1-L2) mCherry ✓ | 6 total

OLIGO(S): 086 (P22) : 0363 (VR) ✓

THERMOCYCLER SETTINGS: 94-2' {94'-15"; 65 ~~-1.5'~~ 15"; 68°C-1.5'}$_{x31}$ 68°C-4'; 4°C

CONTROLS:

POSITIVE:
W/ polymerase ~

NEGATIVE:
w/o polymerase ✓

011 49 2103 2900

CONFIRMATION?
Strong band at ~979 bp
Gel

GEL LANE ORDER:
~~1kb~~ RH1 RH2 RM1 RM2 RL1 RL2
PL1 PL2 PL2 " " "
1P 1N 2P 2N 3P 3N 4P 4N 5P 5N 6P 6N

Gel extract (this will be on multiple gels → ladders w/ matching filenames is essential)

GEL PICTURE / OTHER SPACE: