

**Gradient PCR of - RLuc + addition of hog1 flanking sequence H1**  
**- RLuc + addition of pbs2 flanking sequence P1**  
**- split RLucN + pbs2 flanking sequence P2**

Since we had some trouble with our PCRs and our primers are quite long we decided to try a gradient PCR.

Master Mix  
 GC buffer 250 $\mu$ L  
 dNTPs 25 $\mu$ L  
 Phusion 12,5 $\mu$ L

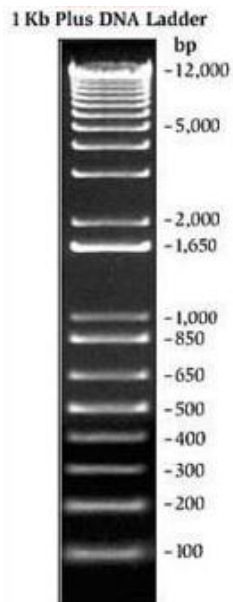
Reaction

	H1	P1	P2
Master Mix	11,5	11,5	11,5
Forward primers 10mM	2,5	2,5	2,5
Reverse primers 10mM	2,5	2,5	2,5
Template pRLuc	0,6	0,6	0,6
Water	33	33	33

Temperature gradient:

H1 64 $\rightarrow$ 72 $^{\circ}$ C  
 P1, P2 55 $\rightarrow$ 63 $^{\circ}$ C

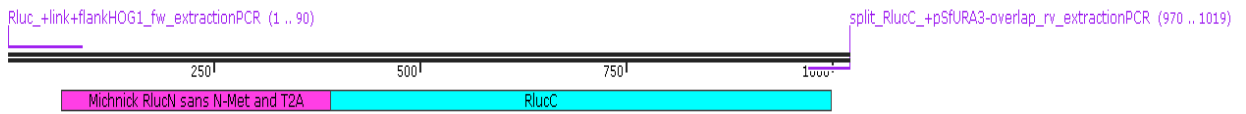
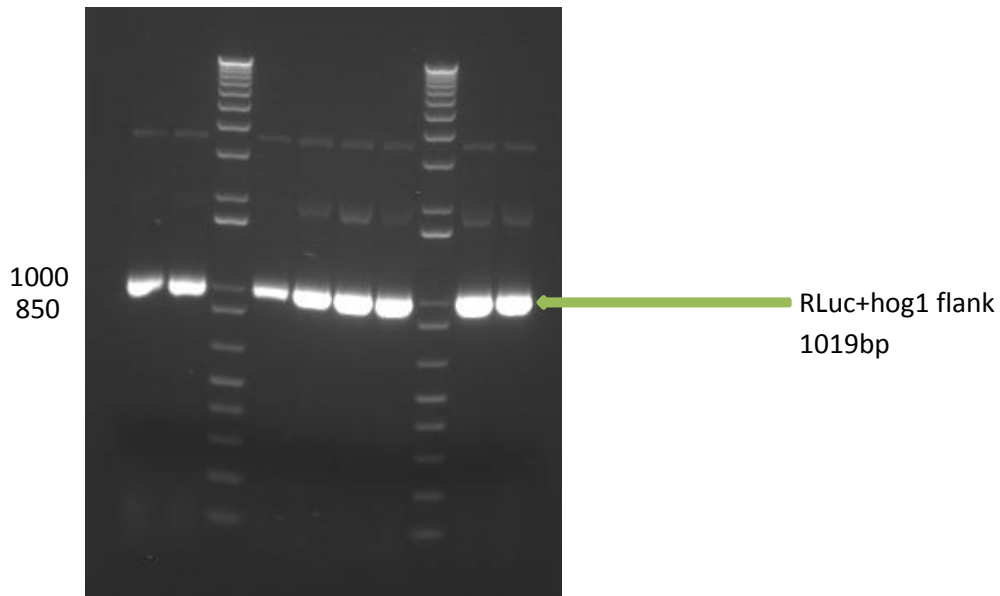
We also increase the extension time: 30seconds $\rightarrow$ 1min30seconds!



We used the Invitrogen 1kb Plus DNA Ladder. In each well of a 1% agarose gel, we loaded 8.8ul Nuclease free Water, 2ul of PCR products or Ladder and 1.2ul 10X Loading Blue Dye. The electrophoresis ran 40 minutes at 120V.

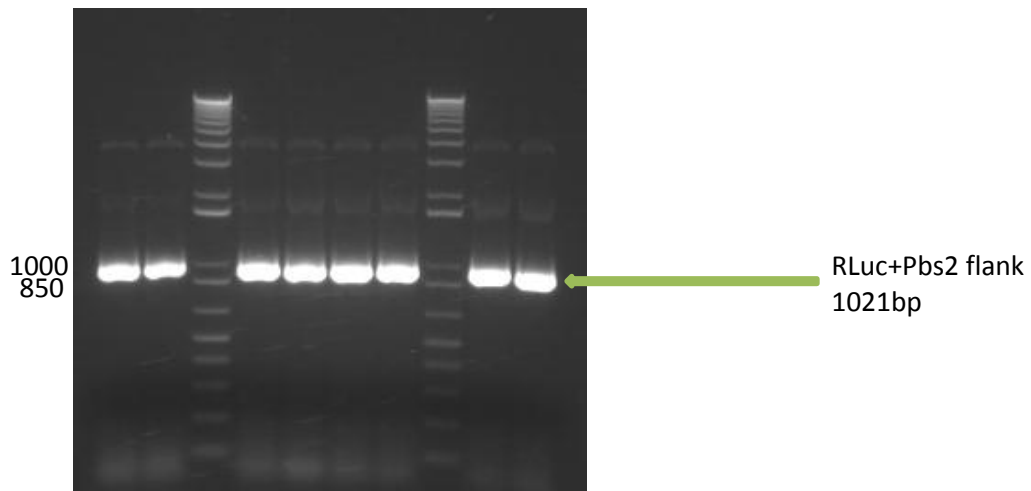
Results:

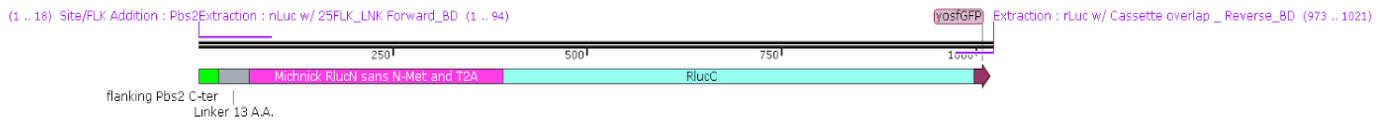
H1



RLuc+hog1flank  
1019 bp

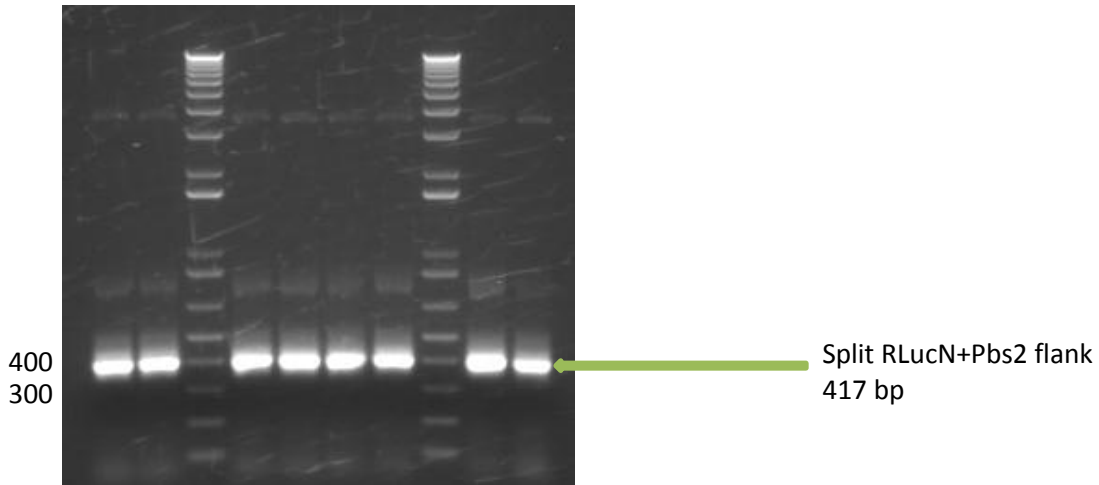
P1





P1\_rLuc-PCR-Intermediate  
1021 bp

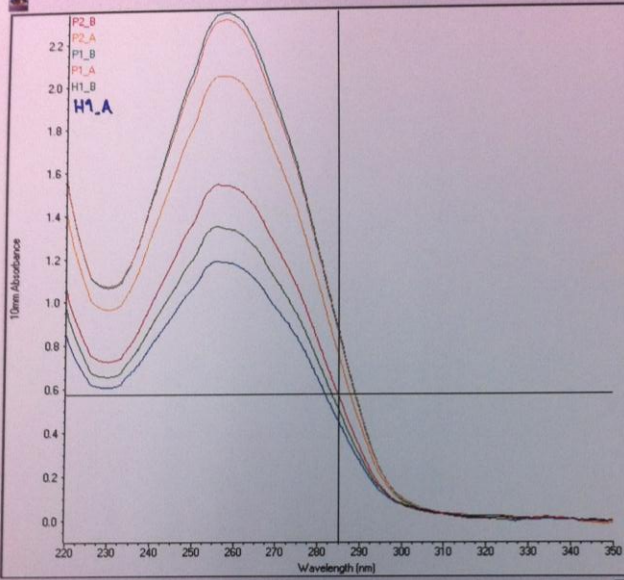
P2



P2\_nLucPCR-intermediate  
417 bp

We purified two PCR samples for each experiment and measured their concentration and purity with the Nanodrop.

#	Sample ID	User name	Date and Time	Nucleic Acid Conc.	Unit	A260	A280	260/280	260/230	Sample Ty
1	H1_A	tp-ssv	12.08.2014 10:10	58.7	ng/ul	1.174	0.652	1.80	1.94	DNA
2	H1_B	tp-ssv	12.08.2014 10:12	66.4	ng/ul	1.329	0.732	1.82	2.04	DNA
3	P1_A	tp-ssv	12.08.2014 10:13	114.7	ng/ul	2.295	1.245	1.84	2.15	DNA
4	P1_B	tp-ssv	12.08.2014 10:15	116.4	ng/ul	2.328	1.261	1.85	2.19	DNA
5	P2_A	tp-ssv	12.08.2014 10:16	101.8	ng/ul	2.037	1.100	1.85	2.12	DNA
6	P2_B	tp-ssv	12.08.2014 10:17	76.4	ng/ul	1.527	0.825	1.85	2.12	DNA



Sample ID: P2\_B Pedestal  
Type: DNA 50.00  
Conc: 76.4 ng/μl  
A260 (10 mm path) 1.527  
A280 (10 mm path) 0.825  
260 / 280 1.85  
260 / 230 2.12  
 Baseline correction 340 nm

285nm 0.572Abs

